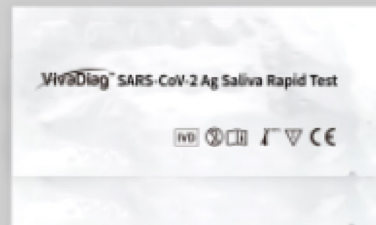


VivaDiag™ SARS-CoV-2 Ag Saliva Rapid Test



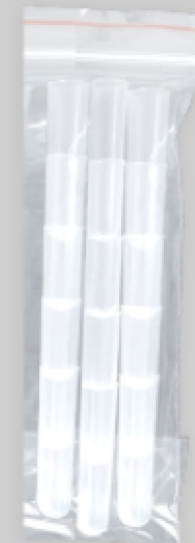
*15 extraction tube tips



*15 test devices



* 15 pipettes



*15 extraction tubes



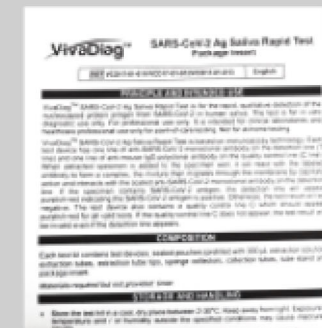
*1 tube stand



*15 extraction solution



* 15 collection cups



*1 package insert



SARS-CoV-2 Ag Saliva Rapid Test Package Insert

REF VCD17-01-011/VCD17-01-012/VCD17-01-013/
VCD17-01-014/VCD17-01-015/VCD17-01-016/VCD17-01-017

English

PRINCIPLE AND INTENDED USE

VivaDiag™ SARS-CoV-2 Ag Saliva Rapid Test is for the rapid, qualitative detection of the nucleocapsid protein antigen from SARS-CoV-2 in human saliva. The test is for *in vitro* diagnostic use only. For professional use only. It is intended for clinical laboratories and healthcare professional use only for point-of-care testing. Not for at-home testing.

VivaDiag™ SARS-CoV-2 Ag Saliva Rapid Test is based on immunoassay technology. Each test device has one line of anti-SARS-CoV-2 monoclonal antibody on the detection line (T line) and one line of anti-mouse IgG polyclonal antibody on the quality control line (C line). When extracted specimen is added to the specimen well, it will react with the labeled antibody to form a complex, the mixture then migrates through the membrane by capillary action and interacts with the coated anti-SARS-CoV-2 monoclonal antibody on the detection line. If the specimen contains SARS-CoV-2 antigen, the detection line will appear purplish-red indicating the SARS-CoV-2 antigen is positive. Otherwise, the test result will be negative. The test device also contains a quality control line C which should appear purplish-red for all valid tests. If the quality control line C does not appear, the test result will be invalid even if the detection line appears.

COMPOSITION

Each test kit contains test devices, sealed pouches (prefilled with 300 µL extraction solution), extraction tubes, extraction tube tips, collection cups, pipettes, tube stand and package insert.

Materials required but not provided: timer.

STORAGE AND HANDLING

- Store the test kit in a cool, dry place between 2-30°C. Keep away from light. Exposure to temperature and / or humidity outside the specified conditions may cause inaccurate results.
- Do not freeze or refrigerate. Use the test kit at temperatures between 15-30°C.
- Use the test kit between 10-90% humidity.
- Do not use the test kit beyond the expiration date (printed on the foil pouch and box).

Note: All expiration dates are printed in Year-Month-Day format. 2022-06-18 indicates June 18, 2022.

WARNINGS, PRECAUTIONS AND LIMITATIONS

- Results from SARS-CoV-2 antigen testing should not be used as the sole basis to diagnose or exclude SARS-CoV-2 infection or to inform infection status.
- Negative results do not rule out SARS-CoV-2 infection, particularly in those who have been in contact with the virus. Follow-up testing with a molecular diagnostic and / or CT should be considered to rule out infection in these individuals.
- Positive results may be due to present infection with SARS-coronavirus strains, see "cross-reactivity" for details. Follow-up testing with a molecular diagnostic and / or CT should be considered to confirm the testing result.
- For *in vitro* diagnostic use only.
- Not for at-home testing.
- Further molecular diagnostic and / or CT is recommended to identify the actual physical situation.
- Do not open the foil pouch of the test device exposing it to the ambient environment until the test device is ready for immediate use.
- Do not use any damaged test device or material.
- Do not reuse the test device.
- Handle extraction solution with caution, do not contact with eyes or skin. If spilled on eyes or skin, wash thoroughly with water.
- Do not use test kit beyond the expiration date.
- Specific training or guidance is recommended if operators are not experienced with specimen collection and handling procedures.
- Only use saliva as specimen. Follow the package insert to obtain accurate results.
- Wear protective gears such as laboratory coats, disposable gloves, and eye protection when specimens are collected and evaluated.
- Wash hands thoroughly after handling.
- All parts of kit are considered biohazardous and can potentially transmit infectious diseases from blood borne pathogens, even after you have performed cleaning and disinfection. Follow proper precautions and all local regulations when disposing of the used test kits.

SPECIMEN COLLECTION AND HANDLING

Please instruct the person who need to test to not place anything in the mouth for at least 10 minutes prior to collection.

1) Specimen collection

Step 1: Remove a collection cup from the box.

Step 2: Spit the saliva into the collection cup, and screw the cap of collection cup.



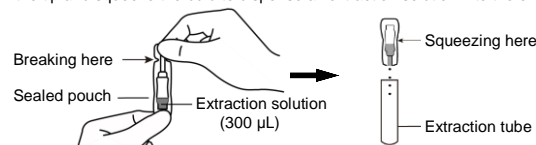
2) Specimen handling

Freshly collected specimens should be tested as soon as possible. It is essential that correct specimen collection and preparation methods are followed.

TEST PROCEDURE

Allow the Test Devices and Extraction Solution to equilibrate to 15-30°C prior to testing.

- Hold the sealed pouch vertically and let all extraction solution flow into the bulb. Break the tip and squeeze the bulb to dispense all extraction solution into the extraction tube.



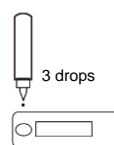
- Collect specimen refer to **Specimen Collection**.
- Use pipette to apply 5 drops of the specimen into the extraction tube, and shake the tube 10 times.



- Put on the tube tip.



- Take out a test device from sealed foil pouch and put it on a clean and level surface.
- Apply 3 drops of the extracted specimen into the specimen well. Please avoid bubbles during applying.



- Read the test result at 15 minutes. Don't read the result after 20 minutes.



15 min



Note:

- Do not interchange or mix extraction solution from different lots.
- Handle extraction solution with caution, do not contact with eyes or skin. If spilled on eyes or skin, wash thoroughly with water.
- Please follow local regulations to handle the used materials.

INTERPRETATION OF TEST RESULTS

1. Positive Result:

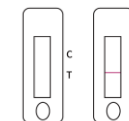
Both the quality control line C and the detection line T appear.

2. Negative Result:

Only the quality control line C appears, with no other line appearing on the detection line.

3. Invalid Result:

Quality control line C fails to appear indicating the test is invalid, no matter if the detection line appears or not. Collect a new specimen and perform another test with a new test device.



Positive: Both detection line (T) and quality control line (C) appear purplish-red in the detection area.

Negative: Only the quality control line (C) appears in the detection area.

Invalid: No purplish-red quality control line (C) appears in the detection areas no matter the detection line (T) is colored or not.

QUALITY CONTROL

Internal procedural controls are included in the test. A colored line appearing in the control region (C) is the internal procedural control. It confirms sufficient specimen volume and correct procedural technique. Control standards are not supplied with this kit; however, it is recommended that positive and negative controls be tested as a good laboratory practice to confirm the test procedure and to verify proper test performance.

PERFORMANCE

1. Limit of Detection

The LOD for the VivaDiag™ SARS-CoV-2 Ag Saliva Rapid Test was established using dilutions of an inactivated virus culture. The starting material was supplied at a concentration of 1.51×10^6 TCID₅₀/mL. Studies were designed to estimate the LOD of the assay using saliva specimens, the starting material was spiked into a volume of pooled human saliva matrix obtained from healthy donors and confirmed negative for SARS-CoV-2 to obtain a series of different concentrations.

SARS-CoV-2 Titer	1.51×10^6 TCID ₅₀ /mL							
Dilution	1/10	1/100	1/1000	1/2500	1/5000	1/10000	1/20000	1/40000
Concentration in Dilution tested (TCID ₅₀ /mL)	1.51×10^5	1.51×10^4	1.51×10^3	6.04×10^2	3.02×10^2	1.51×10^2	75.5	37.8
Detection rates of 5 replicates	100% (5/5)	100% (5/5)	100% (5/5)	100% (5/5)	100% (5/5)	100% (5/5)	100% (5/5)	80% (4/5)
Detection rates of 20 replicates near cut-off	NA	NA	NA	NA	100% (20/20)	100% (20/20)	95% (19/20)	75% (15/20)
Lowest Concentration with Uniform Positivity per Analyte	75.5 TCID ₅₀ /mL							
Limit of detection (LoD) per inactivated Virus Culture	75.5 TCID ₅₀ /mL							

2. Clinical Sensitivity/Clinical Specificity

A total of 603 specimens were tested using the VivaDiag™ SARS-CoV-2 Ag Saliva Rapid Test. These specimens were obtained by saliva from symptomatic patients. The performance of the VivaDiag™ SARS-CoV-2 Ag Saliva Rapid Test was compared to a commercialized molecular assay

Table Summary of sensitivity/specificity of the Ag Saliva Rapid Test compared to PCR.

VivaDiag™ SARS-CoV-2 Ag Saliva Rapid Test	PCR		
	Positive	Negative	Total
Positive	169	0	169
Negative	3	431	434
Total	172	431	603

Sensitivity	98.26% (169/172, 95%CI, 95%~99.41%)
Specificity	100% (431/431 95%CI, 99.12%~100%)
Accuracy	99.50% (600/603 95%CI, 98.55%~99.83%)

The VivaDiag™ SARS-CoV-2 Ag Saliva Rapid Test showed a clinical sensitivity of 98.26%.
The VivaDiag™ SARS-CoV-2 Ag Saliva Rapid Test showed a clinical specificity of 100%.
The VivaDiag™ SARS-CoV-2 Ag Saliva Rapid Test showed a clinical accuracy of 99.50%.

CROSS-REACTIVITY

1. Cross-Reactivity: there was no cross-reaction with potential cross-reactive substances except SARS-coronavirus.

1) cross-reaction with SARS-coronavirus.

Virus	Strain	Concentration
SARS-coronavirus	Urbani	1X10 ⁶ PFU/mL

2) no cross-reaction with potential cross-reactive substances

Virus/Bacteria/Parasite	Strain	Concentration Range
Influenza A	H1N1	1X10 ⁴ ~1X10 ⁶ TCID ₅₀ /mL
	H3N2	
	H5N1	
	H7N9	
Influenza B	NA	
Adenovirus	Type1	
	Type2	
	Type3	
	Type5	
	Type7	
Respiratory syncytial virus	Type A	
	Type B	
Coronavirus	229E	
	OC43	
	NL63	
MERS-Coronavirus	Florida/USA-2_Saudi Arabia.2014	
Parainfluenza virus	Type1	
	Type2	
	Type3	
	Type4	
Rhinovirus A16	N/A	
Legionella pneumophila	Bloomington-2	1X10 ⁵ cells/mL
	82A3105	
Mycobacterium tuberculosis	K	
	Erdman	
	HN878	
	CDC1551	
Streptococcus pneumonia	H37Rv	
	475298	
	[Maryland(D1)6B-17]	
	178[Poland23F-16]	
	262[CIP 104340]	
Streptococcus pyrogens	Slovakia14-10 [29055]	
	Typing stain T1	
Mycoplasma pneumoniae	Mutant22	
	FH strain of Eaton Agent	
	M129-B7	

2. Endogenous/Exogenous Interference Substances Studies: there was no interference for potential interfering substances listed below.

Potential Interfering Substance	Concentration	Results	Viral Strain Culture (in multiples of LoD)	Results
Anti-viral drugs	Zanamivir (Influenza)	5mg/mL	NEG	POS
	Oseltamivir (Influenza)	10mg/mL	NEG	POS
	Artemether-lumefantrine (Malaria)	50uM	NEG	POS
	Doxycycline hyclate (Malaria)	70uM	NEG	POS
	Quinine (Malaria)	150uM	NEG	POS
	Lamivudine (Retroviral medication)	1mg/mL	NEG	POS
	Ribavirin (HCV)	1mg/mL	NEG	POS
Respiratory Specimens	Dacatasvir (HCV)	1mg/mL	NEG	POS
	Mucin: bovine submaxillary gland, type I-S	100ug/mL	NEG	POS
	Blood (human), EDTA anticoagulated	5% (v/v)	NEG	POS
Nasal sprays or drops	Biotin	100ug/mL	NEG	POS
	Neo-Synephrine (Phenylephrine)	10% (v/v)	NEG	POS
	Afrin Nasal Spray (Oxymetazoline)	10% (v/v)	NEG	POS
	Saline Nasal Spray	10% (v/v)	NEG	POS
Homeopathic allergy relief medicine	Homeopathic Zicam Allergy Relief Nasal Gel	5% (v/v)	NEG	POS
	Sodium Cromoglycate	20mg/mL	NEG	POS
	Olopatadine Hydrochloride	10mg/mL	NEG	POS
Anti-inflammatory medication	Acetaminophen	199uM	NEG	POS
	Acetylsalicylic acid	3.62mM	NEG	POS
	Ibuprofen	2.425mM	NEG	POS
Antibiotic	Mupirocin	10mg/mL	NEG	POS
	Tobramycin	5ug/mL	NEG	POS
	Erythromycin	81.6uM	NEG	POS
	Ciprofloxacin	30.2uM	NEG	POS

3. High-dose Hook Effect: cultured SARS-CoV-2 virus was spiked into specimen.
No hook-effect was observed at 1.51X10⁵ TCID₅₀/mL of cultured SARS-COV-2 virus.

Specimen Type	Dilution	Concentration (TCID ₅₀ /ml)	Result
SARS-CoV-2 Inactivated virus cultured	NEAT	1.51x 10 ⁶	POS
	1/10	1.51x 10 ⁵	POS
	1/100	1.51x10 ⁴	POS
	1/1000	1.51x10 ³	POS
	1/2500	6.04x10 ²	POS
	1/5000	3.02x10 ²	POS
	1/10000	1.51x10 ²	POS
	1/20000	75.5	POS

	1/40000	37.8	NEG
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POS: positive

NEG: negative

REFERENCES

1. Coronaviridae Study Group of the International Committee on Taxonomy of Viruses. The species severe acute respiratory syndrome-related coronavirus: classifying 2019-nCoV and naming it SARS-CoV-2 [J]. Nature Microbiology, 5, 536-544 (2020).
2. Perlman, S. Netland, J. Coronaviruses post-SARS: update on replication and pathogenesis. Nature Reviews Microbiology 7, 439-450, doi: 10.1038/nrmicro2147 (2009).
3. Lauer SA, Grantz KH, Bi Q, et al. The Incubation Period of Coronavirus Disease 2019 (COVID-19) From Publicly Reported Confirmed Cases: Estimation and Application. Ann Intern Med. 2020; 172(9): 577-582. doi: 10.7326/M20-0504.
4. McCarron, MM, et al. "Detection of Phencyclidine Usage by Radioimmunoassay of Saliva," J Anal Tox. 1984 Sep-Oct.; 8 (5), pp 197-201.

INDEX OF SYMBOLS

	Consult instructions for use		Use by		Contains sufficient for <n> tests
	For in vitro diagnostic use only		Lot number		Catalog number
	Storage temperature limitations		Manufacturer		Do not reuse
	Authorized Representative				

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